

**Title:** Case studies in neuroscience: Subcortical origins of the frequency-following response

**Running title:** Subcortical origins of the FFR

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**Abstract:** The auditory frequency-following response (FFR) reflects synchronized and phase-locked activity along the auditory pathway in response to sound. While historically thought to reflect subcortical activity, recent evidence suggests an auditory cortex contribution as well. Here we present electrophysiological evidence for the FFR's origins from two cases: a patient with bilateral auditory cortex lesions and a patient with auditory neuropathy, a condition of subcortical origin. The patient with auditory cortex lesions had robust and replicable FFRs, but no cortical responses. In contrast, the patient with auditory neuropathy had no FFR despite robust and replicable cortical responses. This double dissociation shows that subcortical synchrony is necessary and sufficient to generate an FFR.

**Keywords:** Frequency-following response, auditory cortex lesions, auditory neuropathy, electrophysiology, auditory evoked potentials

## Introduction

The FFR has emerged in neuroscience as a means to understand auditory behaviors such as speech and music perception, real-world listening, and experience-dependent plasticity in auditory processing. FFRs have long been thought to originate primarily in subcortical auditory nuclei, with the inferior colliculus of the auditory midbrain serving a chief role; indeed FFRs recorded directly from the midbrain strongly resemble those recorded at the scalp (White-Schwoch et al. 2017) and cooling the inferior colliculus eliminates FFRs (Smith et al. 1975). This hypothesis is also supported by evidence in humans. Sohmer and Pratt (1977) showed that FFRs were absent in 9 patients with brainstem lesions. Bidelman (2015, 2018) recorded FFRs with high-density, multichannel EEG and used source modeling to show that FFRs reflect sustained auditory midbrain activity.

Recent MEG evidence indicates a role for the right hemisphere auditory cortex in generating FFRs to the fundamental frequency ( $F_0$ ), along with contributions from the brainstem, midbrain, and thalamus (Coffey et al. 2016) (see (Zhao and Kuhl 2018) for additional MEG evidence pointing to subcortical sources of the FFR). Some authors have suggested these MEG results should be considered when interpreting EEG FFRs (Coffey et al. 2016, 2017; Holmes and Herrmann 2017; but see Bidelman 2018), the prevailing approach for recording FFRs. This raises the question: is the auditory midbrain necessary and sufficient to generate an FFR?

To provide clarity to the origins of the scalp-recorded FFR, we conducted a dual case study. Specifically, we conducted an electrophysiological test battery in two patients who provide rare opportunities to test multiple hypotheses about the origins of the FFR.

## Materials and Methods

This study compares auditory-neurophysiological responses in two patients: NR, who has bilateral auditory cortex lesions, and IT, who has auditory neuropathy. Details on their case histories are presented in the results section, but both have essentially normal cochlear function (see Figs. 1 and 2). Both patients provided written consent to participate in the project, and procedures were approved by the Institutional Review Board of Northwestern University.

Auditory brainstem responses (ABRs), FFRs, and cortical potentials were recorded in both patients.

The stimulus for ABRs was a 100  $\mu$ s click presented at 80 dB SPL and 31.1 Hz and recorded with a Bio-Logic NavPro system. Clicks were presented in rarefaction (and, for IT, also condensation) polarity, and 2,000 artifact-free responses were recorded. Responses were averaged from 100-1,500 Hz online. We also have records of an ABR latency-intensity function for NR (see Fig. 1c), which used rarefaction clicks at 13.3 Hz with similar filtering, and also on a Bio-Logic NavPro.

The stimulus for FFRs and cortical potentials was a 170-ms [da] used in previous studies (Anderson et al. 2012). In addition to the [da] collected in quiet, we presented it in background noise, a six-talker babble track (timed to avoid phase synchrony with the [da]). We wanted a second, more challenging stimulus condition to compare to responses in quiet. This would let us test both whether responses are absent or present, and test for changes in response morphology as a function of listening condition (cf. White-Schwoch et al. 2015). We chose noise given evidence that patients with neuropathy have significant difficulties understanding speech in noise (Kraus et al. 2000).

For FFRs, the [da] was presented at 3.9 Hz at 80 dB SPL in alternating polarities; 6000 artifact-free responses were collected in the quiet and background noise conditions from Cz, filtered from 70-2000 Hz, and epoched from 0-210 ms re stimulus onset. The SNR for the noise condition was +10 dB. NR's FFRs were recorded in a Neuroscan SynAmps system and IT's FFRs were recorded in a Biosemi Active2 system.

Spectra were computed via fast Fourier transforms. For cortical responses, the [da] was presented at 1 Hz at 80 dB SPL in alternating polarity; 500 artifact-free responses were collected in each condition from Cz, filtered from 1-40 Hz, and epoched from -100-500 ms re stimulus onset. Similar to the FFR, the SNR for the noise condition was +10 dB. Both responses were recorded in a Neuroscan SynAmps system; NR's responses were recorded with a 32-channel cap and IT's with a single channel at Cz.

## Results

The first case, NR, is a 28-year-old male with bilateral auditory cortex lesions following treatment for acute lymphoblastic leukemia. His treatment included a breathing tube that was dislodged, causing hypoxia, followed by a medically induced coma, multiple rounds of chemotherapy, and a bone marrow transplant. Following his treatment he experienced hearing difficulties. His medical team conducted an MRI and the radiologist's report identified bilateral temporal lobe lesions.

NR is cortically deaf: he lacks awareness of sounds and cannot understand speech or music. He also exhibits signs of mild visual agnosia. Cognitive abilities were otherwise normal. Due to his poor ability to detect sounds, NR could not complete a behavioral audiogram. But, NR had normal distortion product otoacoustic emissions (DPOAEs; see Fig. 1a for a DP-gram)

and auditory brainstem responses (ABRs; Fig. 1b/1c), indicating an origin central to the auditory midbrain for his deafness. Consistent with the diagnosis of cortical deafness, NR had absent cortical auditory evoked potentials (Fig. P1/N1; Fig. 2e).

Yet, NR had robust and highly replicable FFRs (Fig. 1a). Latencies and amplitudes were within normal limits and similar to healthy, age-matched controls (Anderson et al. 2012). NR had robust responses to the fundamental frequency in both quiet and background noise (Fig. 1b), which is noteworthy because MEG source modeling ascribes the FFR to the 100-Hz F0 of this stimulus, at least in part, to the auditory cortex (Coffey et al. 2016). NR's robust response to the F0 suggests that multiple sources can contribute to the F0 component of the FFR, consistent with MEG source analysis (Coffey et al. 2016)—and NR's response shows that cortex is not necessary for generating low frequency components of the FFR. NR's case is consistent with studies in cats that were lesioned in either the IC or auditory cortex, and showed that IC was necessary and sufficient to generate FFRs (Kiren et al. 1994).

The second case, IT, is a 41-year-old woman with auditory neuropathy, meaning she has intact outer hair cell function in the cochlea but an absent auditory brainstem response due to dyssynchronous subcortical firing (Starr et al. 1996). IT has tremendous difficulty understanding speech in adverse listening conditions, such as noisy environments, but has normal speech perception in quiet. She also reports some difficulty with sound awareness, albeit not to the extent of NR, and difficulty understanding unfamiliar accents. IT had a normal audiogram (Fig. 2a) and normal DPOAEs (Fig. 2, inset) but an absent ABR bilaterally (Fig. 2b/c). Additionally, and consistent with her absent ABR, IT had no FFR (Fig. 3b, 3d).

Yet, she had robust and highly replicable cortical responses (Fig. 3f). Her response in quiet was somewhat larger than typically observed in adults (cf. Cunningham et al. 2000), albeit later, and her response in noise was particularly delayed. This may reflect cortical-subcortical interactions, particularly for coding speech features in noise. Her cortical response is also evocative of that found in older adults, where larger responses are thought to reflect compensatory mechanisms (Bidelman et al. 2014; Roque et al. 2019). Moreover, her response likely lacks normal brainstem inhibitory processes. It is noteworthy that her response in noise is especially delayed given her difficulties understanding speech in noise. Cortical-subcortical interactions may partially drive cortical responses to speech in noise. Still, overall, her case is consistent with the notion that a functioning auditory cortex is not sufficient to generate an FFR.

## Discussion

Together, NR and IT show a double dissociation: auditory cortex is necessary for cortical responses but unnecessary for FFRs. Highly precise subcortical synchrony is necessary for FFRs but unnecessary for cortical responses. The auditory midbrain and cortex may specialize to fill distinct roles in auditory perception. If the cortex does not receive synchronous input from the midbrain, the signal will be temporally smeared such that the cortex will not be able to reconstruct it. Indeed, listeners with neuropathy exhibit profound deficits with fast temporal processing (Kraus et al. 2000; Starr et al. 1996; Zeng et al. 1999). In contrast, NR's case shows that this fine temporal resolution can be intact but is insufficient for speech communication or music perception.

One drawback of this study is that we do not have detailed anatomical information about NR's lesions. We have the radiologist's report, which indicates bilateral temporal lobe lesions but does not provide detailed information about the possibility of residual auditory fields. We note the complete absence of a cortical evoked potential, which is consistent with auditory cortex lesions (Özdamar et al. 1982). Nevertheless, with our current data we cannot rule out the possibility of residual cortical contributions to his FFR, although we think it unlikely that such a contribution would contribute to the FFR but *not* generate any cortical evoked potential (i.e. consist only of 100 Hz phaselocking).

In IT's case we similarly do not have a precise localization of the "site of lesion" – her neuropathy may be pre- and/or post-synaptic. We note evidence of an MLR visible in her FFR tracings (in the 25-75 ms range; see Fig. 3b) consistent with functioning thalamic and corticothalamic pathways (Kraus et al. 1982, 1988) and reinforces evidence of a subcortical neuropathy. Still, a number of subcortical insults and lesions can produce the auditory neuropathy phenotype (Kraus et al. 1984; Starr et al. 1996, 2003).

This is neither to gainsay cortical contributions to healthy subjects' FFRs, nor to undermine the strong, long-term influence of cortical processing on subcortical auditory function. As with any case study, our results depend on rare circumstances that may not generalize to all subjects. Noteworthy, however, is their consistency with early FFR work in humans (Sohmer et al. 1977), recent source modeling work (Bidelman 2015, 2018), and lesion studies in animals (Kiren et al. 1994). When considered in tandem with previous work, these case studies show that accurate and synchronous encoding of the auditory midbrain is necessary and sufficient to generate an electrophysiological FFR.



**Acknowledgments:** We are grateful to NR, IT, and their families for their participation in research. We also thank R. Zatorre for his input, A. Starr, and colleagues in the Auditory Neuroscience Laboratory. Supported by the Knowles Hearing Center.

**Competing Interests:** The authors declare no competing interests.

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## Figure Captions

**Figure 1:** Results from NR's audiology testing. (A) Distortion product otoacoustic emission results for the right (red) and left (blue) ears from 516-10,008 Hz. Plotted is the SNR of the emission (distortion product minus noise floor, DP-NF). Overall NR has normal cochlear function, with a possible notch noted in the right ear. He may have a mild sensorineural hearing loss from chemotherapy. (B) ABRs collected in the Auditory Neuroscience Laboratory in response to a rarefaction click presented at 31.1 Hz and 80 dB SPL. Thin lines show two runs of the click ABR with the bold line showing their average. While the morphology is somewhat idiosyncratic, the response is reliable. (C) ABR latency-intensity study provided to us by NR's audiologist. Both ears show normal responses to changes in intensity (note this click was delivered at 13.3 Hz).

**Figure 2:** Results from IT's audiology testing. (A) Audiogram shows normal hearing thresholds. Right ear is shown in red, left ear in blue, and bone-conduction testing in grey. The inset shows DPOAEs in the right and left ears, which are robust. (B/C) ABRs were elicited at 31.1 Hz and 80 dB SPL in rarefaction (bright colors) and condensation (duller colors) in the right and left ears, respectively. Consistent with her diagnosis of auditory neuropathy, IT had absent ABRs.

**Figure 3:** Subcortical and cortical responses are shown for both NR and IT. Each panel is organized analogously. The top two rows shows FFRs to "da" in quiet (black) and noise (purple) in the time and frequency domains, respectively. NR has robust responses in both conditions, including to the F0 at 100 Hz, which were within normal limits (Anderson et al. 2012). IT has no FFR. The bottom row shows cortical responses to the same stimuli. NR has no cortical response but IT has a robust P1/N1 response both in quiet and noise.





